

Manual

Genomic Mini AX Plant Spin

Increased efficiency kit for genomic DNA purification from plant tissue.

catalog#	size
050-100S	100 isolations

For research use only.

Guarantee

A&A Biotechnology provides a guarantee on this product.

The company does not guarantee the correct performance of this kit in the event of:

- not adhering to the supplied protocol
- use of not recommended equipment or materials
- use of other reagents than recommended or which are not a component of the product
- use of expired or improperly stored product or its components

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Contents

component	100 isolations	storage
Mini AX Spin columns	100 pcs	2-8°C
2 ml tubes	200 pcs	15-25 ℃
LS lysis suspension	100 ml	15-25 ℃
W1 first wash solution	70 ml	15-25 ℃
W2 second wash solution	60 ml	15-25 ℃
E elution buffer (without EDTA)	20 ml	2-8°C
N neutralizing buffer	1 ml	15-25 ℃
T solution	400 μΙ	2-8°C
Proteinase K	2 x 1.1 ml	2-8°C

The binding capacity of the column is 15 $\mu\text{g}.$

Additional equipment and reagents

Necessary

- 1.5 ml sterile Eppendorf tubes
- Incubator or thermoblock set to 50 °C
- Vortex
- Microcentrifuge

Optional

RNAse (cat. # 1006-10, 1006-50)

Important information

• E elution buffer loses activity upon prolonged contact with air. Always close the E elution buffer vial tightly directly after use. Store E elution buffer at 2-8 °C.

Isolation protocol

1.	Transfer up to 20 mg of dried, powdered plant material or up to 100 mg fresh /frozen cut plant tissue to a 1,5 ml Eppendorf tube (not included).
2.	Note: LS lysis suspension should be mixed by inverting the tubes before use. Add 900 μ l of LS lysis suspension and 20 μ l of proteinase K.
3.	Vortex the sample and incubate for 10 min at 50 °C. Vortex the sample from time to time. The incubation step can be performed in Eppendorf Thermomixer or analogous equipment at 1400 RPM and 50 °C. RNA digestion (optional): add 5 µl of RNAse (10 mg/ml solution) (not included). Mix and incubate for 5 min at room temp.
4.	Intensively vortex the sample for 2 min at 1000-1400 RPM . This is the key step for efficiency of DNA isolation.
5.	Centrifuge for 5 min at 14 000 x g . The DNA pellet should be visible at the bottom of the tube. It is a mixture of non-lysed fragments of sample material and particles of the LS lysis suspension.
6.	Apply 600 µI of supernatant onto the Mini AX Spin column placed inside a 2 ml tube.
7.	Centrifuge for 30-60 s at 8 000 x g.
8.	Transfer the Mini AX Spin column to a new 2 ml tube (included).
9.	Add $600\mu l$ of $W1$ first wash solution. Centrifuge for $30\text{-}60s$ at $8000xg$.
10.	Transfer the Mini AX Spin column to a new 2 ml tube (included).
11.	Add 500 μI of W2 second wash solution.

	Centrifuge for 30-60 s at 14 000-21 000 x g.
12.	Prepare a 1.5 ml elution tube (not included) and add 5 μ l of N neutralizing buffer. DNA neutralization - page 6.
13.	Transfer the Mini AX Spin column to the prepared elution tube.
14.	Before using E buffer, it is recommended to do a functionality test - page 6. Apply 100-150 µl of E elution buffer onto the Mini AX Spin column. Keep for 2 min at room temp. E elution buffer loses activity upon prolonged contact with air. Always close the E elution buffer vial tightly directly after use. Store E elution buffer at 2-8 °C.
15.	Centrifuge for 30-60 s at 14 000-21 000 x g.
16.	Remove the Mini AX Spin column. Close the tube with purified DNA.

DNA neutralization

E elution buffer is strongly alkaline and may cause DNA degradation upon freezing. Thus it is necessary to use a N neutralizing buffer. We recommend adding the N neutralizing buffer to the elution tube before the elution step. If the N neutralizing buffer was not added before the elution step it can be added directly before freezing DNA samples. The use of N neutralizing buffer enables secure DNA storage conditions at 10 mM TrisHCl, pH 8.5.

E buffer functionality test

E buffer has a critical influence on DNA elution efficiency and thus overall DNA purification yield. The kit contains T solution which enables testing of the E buffer correct functionality.

Typically it is suggested to perform such a test in the following cases:

- E buffer was not used for a long period of time (at least 2 months).
- E buffer was stored at room temp. for a long period of time (at least 2 weeks).
- E buffer vial was not closed tightly.

Procedure:

Transfer $20\,\mu$ I of E buffer to PCR tubes; add $2\,\mu$ I of T solution; mix the sample, wait $2\,\mu$ min. Compare the mixture color with the reference color guide.



Safety information



DANGER

Proteinase K

H315 Causes skin irritation.

H319 Causes serious eye irritation.

H334 May cause allergy or asthma symptoms or breathing difficulties if inhaled.

H335 May cause respiratory irritation.

P261 Avoid breathing dust.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,

if present and easy to do. Continue rinsing.

P342+P311 If experiencing respiratory symptoms call a Poison Center or doctor/physician.

LS lysis suspension



WARNING

H302 Harmful if swallowed.

H315 Causes skin irritation.

H319 Causes serious eye irritation.

 $P305 + P351 + P338 \ If \ in \ eyes: rinse \ cautiously \ with \ water for \ several \ minutes. \ Remove \ contact \ lenses,$

if present and easy to do. Continue rinsing.





DANGER

H225 Highly flammable liquid and vapor.

H319 Causes serious eye irritation.

W1 first wash solution

H336 May cause drowsiness or dizziness.

P210 Keep away from heat, sparks, open flames, hot surfaces. No smoking.

P261 Avoid breathing vapors.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,

if present and easy to do. Continue rinsing.



DANGER

E elution buffer

H314 Causes severe skin burns and eye damage.

P280 Wear protective gloves/ protective clothing/ eye protection/ face protection.
P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,

if present and easy to do. Continue rinsing.

P310 Immediately call a Poison Center or doctor/physician.



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