



A&A BIOTECHNOLOGY
innovating life science

Genomic Mini

Versatile kit for genomic DNA purification from various sources.

version 0517

50 isolations, 250 isolations

Cat. # 116-50, 116-250

Kit for DNA isolations from z tissues, bacteria, cell culture.
The binding capacity of the DNA purification column is 20 µg of DNA.

For R&D use only

Kit Contents

Component	50 isolations	250 isolations	Store at
Minicolumns	50 pcs	250 pcs	Room Temp.
2 ml tubes	50 pcs	250 pcs	Room Temp.
A1 wash solution	50 ml	250 ml	Room Temp.
LT lysis solution Note: If LT solution contains the precipitate, heat it to 40 °C until dissolution of complete precipitate.	15 ml	60 ml	Room Temp.
Tris buffer (10 mM, pH 8,5)	25 ml	125 ml	Room Temp.
Proteinase K	1.1 ml	5 x 1.1 ml	+4 to +8 °C

Equipment and materials necessary for the DNA isolation that are not included in kit

1. Material for DNA isolation
2. Enzymes (option – depending on type of biological material):
 - Lysozyme – 10 mg/ml, 400 U/μl (cat. # 1005-10, 1005-50)
 - Lysostaphin – 0.4 U/μl (cat. # 1007-400, 1007-2000)
 - Mutanolysin – 10 U/μl (cat. # 1017-5, 1017-10, 1017-50)
3. RNase – 10 mg/ml (cat. # 1006-10, 1006-50) (optional)
4. Sterile water (nuclease free, DEPC treated) (cat. # 003-075, 003-25) (optional)
5. DTT (dithiothreitol) (cat. # 2010-5, 2010-25, 2010-10P) (optional)
6. 1.5 ml Eppendorf tubes
7. Incubator or heatblock set to 37 °C, 50 °C, 70 °C
8. Vortex
9. Microcentrifuge

NOTE:

Before you start working, we recommend cleaning the work surface using LabZAP™ product (cat. # 040-500).

A&A Biotechnology provides one year guarantee on this kit

The company does not guarantee correct performance of this kit in the event of:

- not adhering to the supplied protocol
- not recommended use of equipment and materials
- the use of other reagents than recommended or which are not a component of the kit
- the use of expired or improperly stored reagents and minicolumns

Material preparation

Bacteria

1. Transfer 100 µl of bacterial culture to a 1.5 ml tube (not included).

NOTE: For bacterial culture 200 µl–1 ml volume: centrifuge the sample, discard supernatant, suspend the bacterial pellet in 100 µl of Tris buffer.

2. For Gram(+) bacteria, we recommend using the following enzymes:

for *S.aureus* we recommend using lysostaphin (0.4 U/µl)

(not included, cat # 1007–400, 1007–2000);

add 10 µl of lysostaphin and incubate for 10 min at 37 °C.

for *Streptococcus*, *Lactobacillus*, *Lactococcus*, *Listeria* we recommend using mutanolysin

(10 U /µl) (not included, cat. # 1017–5, 1017–10, 1017–50) or mutanolysin with

lysozyme (not included, cat # 1005–10, 1005–50);

add 5 µl of mutanolysin or 5 µl of mutanolysin and 10 µl of lysozyme.

Mix and incubate for 20 min at 50 °C.

3. Add 200 µl of LT lysis solution and 20 µl of Proteinase K solution.
4. Mix the whole sample by inverting the tube and incubate for 20 min at 37 °C.
RNA digestion (optional): Add 5 µl of RNase (10 mg/ml solution) (not included, cat. # 1006–10, 1006–50). Mix sample and incubate for 5 min at room temp.
5. Follow point 1. of the protocol

Semen

1. Transfer 100 µl of semen to a 1.5 ml tube (not included).
2. Add 10 µl of 1M DTT (not included, cat # 2010–5, 2010–25, 2010–10P).
3. Add 200 µl of LT lysis solution and 20 µl of Proteinase K solution.
4. Mix the sample by inverting the tube and incubate for 20 min at 37 °C.
RNA digestion (optional): Add 5 µl of RNase (10 mg/ml solution) (not included, cat. # 1006–10, 1006–50). Mix sample and incubate for 5 min at room temp.
5. Follow point 1. of the protocol

Cell culture

1. Transfer 1×10^6 of cell culture to a 1.5 ml tube (not included). Centrifuge and discard the supernatant.
2. Suspend the pellet in 100 μ l of Tris buffer.
3. Add 200 μ l of LT lysis solution and 20 μ l of Proteinase K solution.
4. Mix the sample by inverting the tube and incubate for 20 min at 37 °C.
RNA digestion (optional): Add 5 μ l of RNase (10 mg/ml solution) (not included, cat. # 1006-10, 1006-50). Mix sample and incubate for 5 min at room temp.
5. Follow point 1. of the protocol

Fresh tissues

1. Up to 10–15 mg of fragmented tissues transfer to a 1.5 ml tube (not included).
2. Add 100 μ l of Tris buffer, 50 μ l of LT lysis solution and 20 μ l of Proteinase K solution.
3. Mix the sample by vortexing. Incubate at 50 °C up to completely lyse of tissue. Mix the sample from time to time by vortexing.
4. Mix the sample by vigorous vortexing for 20 s.
RNA digestion (optional): Add 5 μ l of RNase (10 mg/ml solution) (not included, cat. # 1006-10, 1006-50). Mix sample and incubate for 5 min at room temp.
5. Add 150 μ l of LT lysis solution and mix the sample.
6. Follow point 1. of the protocol

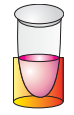
Embedded tissues

1. Transfer the tissue to a 1.5 ml tube (not included).
2. Add Tris buffer, centrifuge and discard supernatant. Repeat several times to remove the fixing liquid.
3. Remove the paraffin from FFPE samples – rinse in xylene (not included), next rinse in ethanol (not included).
4. Follow the fresh tissues of material preparation.

For embedded tissues we recommend Xpure™ FFPE micro (cat. # 091-50) – Kit for genomic DNA purification from tissues preserved in paraffin. Fast deparaffinization without xylene and hexane.

Isolation protocol

1. Set the thermoblock temperature to 70 °C and place in it the tubes with Tris elution buffer (it will be used in point 8. of protocol).



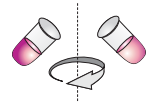
70 °C

2. Incubate the samples for 5 min at 70 °C.



70 °C

3. Mix by vortexing for 20 s and centrifuge for 1 min at 10 000–15 000 RPM.



4. Apply the supernatants onto the minicolumns.

Centrifuge for 1 min at 10 000–15 000 RPM.



5. Add 500 µl of A1 wash solution.

Centrifuge for 1 min at 10 000–15 000 RPM.

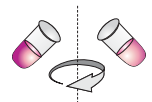


6. Transfer the minicolumns to the new 2 ml tubes (included).

Add 400 µl of A1 wash solution.



7. Centrifuge for 1 min at 10 000–15 000 RPM.



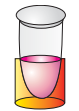
8. Transfer the minicolumns to **the new 1.5 ml** tubes (not included).

Add to the minicolumns **200 µl** of **Tris** buffer (included) or sterile water (not included) heated to **70 °C**.

NOTE: If it is expected the low isolation yield, the elution volume (Tris buffer or water) can be reduced to 100 µl increasing the final DNA concentration.

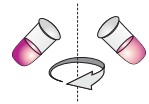


9. Incubate the samples for **2 min** at **room temp.**



**room
temp.**

10. Centrifuge for **1 min** at **10 000–15 000 RPM**.



11. Remove the minicolumns.
Store the purified DNA at **+4 °C** or **-20 °C**.

Related products

Product	Quantity	Cat. #
Xpure Genomic Mini	50 isolations	096-50
Xpure Cell&Tissue	50 isolations	090-50
Genomic Midi AX	20 isolations	895-20
Genomic Midi AX Direct	20 isolations	895-20D
Genomic Maxi AX	10 isolations	995-10
Genomic Maxi AX Direct	10 isolations	995-10D
Plasmid Mini	50 isolations	020-50
	250 isolations	020-250
Gel-Out	50 isolations	023-50
	250 isolations	023-250
Clean-Up	50 isolations	021-50
	250 isolations	021-250
ExTerminator	50 isolations	444-50
	250 isolations	444-250
Swab	25 isolations	025-25
	100 isolations	025-100
Genomic Mini AX Swab&Semen Spin	100 isolations	025-100S

Safety information



DANGER

Proteinase K

H315 Causes skin irritation.

H319 Causes serious eye irritation.

H334 May cause allergy or asthma symptoms or breathing difficulties if inhaled.

H335 May cause respiratory irritation.

P261 Avoid breathing dust.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P342+P311 If experiencing respiratory symptoms call a Poison Center or doctor/physician.



WARNING

LT lysis solution

H302 Harmful if swallowed.

H315 Causes skin irritation.

H319 Causes serious eye irritation.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.



DANGER

A1 wash solution

H225 Highly flammable liquid and vapour.

H319 Causes serious eye irritation.

H336 May cause drowsiness or dizziness.

P210 Keep away from heat, sparks, open flames, hot surfaces. No smoking.

P261 Avoid breathing vapours.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.