

Manual

Genomic Mini AX Bacteria

Increased efficiency kit for genomic DNA purification from bacteria Procedure with DNA precipitation.

catalog #	size
060-60	60 isolations

For research use only.

Guarantee

A&A Biotechnology provides a guarantee on this product.

- The company does not guarantee the correct performance of this kit in the event of:
 - not adhering to the supplied protocol
 - use of not recommended equipment or materials
 - use of other reagents than recommended or which are not a component of the product
 - use of expired or improperly stored product or its components



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Contents

component	060-60	storage
Genomic Mini AX columns	60 pcs	2-8 °C
2 ml tubes	60 pcs	15-25 ℃
BS suspension buffer	7 ml	2-8 °C
LS lysis suspension	60 ml	15-25 ℃
K1 equilibrating solution	55 ml	15-25 ℃
K2 wash solution	190 ml	15-25 ℃
K3 elution solution	70 ml	15-25 ℃
PM precipitation mix	55 ml	15-25 ℃
Tris buffer (10 mM Tris-HCl, pH 8,5)	10 ml	15-25 ℃
Lysozyme	650 µl	-20 °C
Proteinase K	1.3 ml	2-8 °C

The binding capacity of the minicolumn is $20 \ \mu g$.

Additional equipment and reagents

Necessary

- 1.5 ml, 2 ml Eppendorf tubes
- 70% ethanol
- Incubator or thermoblock set to 37 °C, 50 °C
- Vortex
- Microcentrifuge

Optional

- Lysostaphin 15 U/µl (cat. # 1007-3, 1007-15) / Mutanolysin 10 U/µl (cat. # 1017-5, 1017-10)
- RNAse (cat. # 1006-10, 1006-50)
- Sterile water (cat.# 003-075, 003-25)
- TE buffer (cat.# 297-100)

Important notes

• The chromatography purification of DNA can be paused at any time while a sample is loaded onto a column. The purification process can be continued after a 15-hours-long pause with no influence on quality or quantity of purified DNA. During the pause of the DNA purification the 15 ml tube with the column inside has to be closed with the screw cap to avoid membrane desiccation and subsequent DNA loss.

Protocol

1.	Centrifuge 0.2-1 ml of overnight bacterial culture . Discard the supernatant. Suspend the bacterial pellet in 100 μl of BS suspension buffer.
2.	Add 10 µl of Iysozyme . for S <i>aureus</i> we recommend using Iysostaphin (not included); for <i>Streptococcus, Lactobacillus, Lactococcus, Listeria</i> we recommend using mutanolysin recombinant (not included). Mix and incubate for 15 min at 37 °C .
3.	LS lysis suspension should be mixed by inverting the tubes before use. Add 900 μl of LS lysis suspension and 20 μl of proteinase K .
4.	Vortex the sample and incubate for 30 min at 50 °C . Mix the samples by inverting the tubes a few times. The incubation step can be performed in Eppendorf Thermomixer or analogous equipment at 1400 RPM and 50 °C. RNA digestion (optional): add 5 µl of RNAse (10 mg/ml) (not included). Mix and incubate for 5 min at room temp.
5.	During incubation prepare the Genomic Mini AX columns placed inside 15 ml tubes. Apply 800 µl of K1 equilibrating solution. Wait for the solution to flow through the column.
6.	Vortex the sample for 15 s . Centrifuge for 5 min at 10 000-14 000 RPM . The DNA pellet should be visible at the bottom of the tube. It is a mixture of non-lysed fragments of sample material and particles of the LS lysis suspension.
7.	Apply the supernatant onto the equilibrated Genomic Mini AX column. Wait for the lysate to flow through the column. The Genomic Mini AX column works by means of gravity. The flow rate strongly depends directly on the quantity and size of DNA molecules in a sample. High content of high molecular weight DNA decreases the flow rate. DNA amount exceeding 20 µg loaded onto a column may lead to flow stoppage. In such cases the column should be centrifuged in a swing-out rotor for 1 min at 3000-4000 RPM. The centrifugation can be performed after the loading step (point 7) and during the washing step with K2 solution (point 8 and 9) or elution step with K3 solution (point 10).
8.	Add 1.5 ml of K2 wash solution. Wait for the solution to flow through the column.

9.	Add again 1.5 ml of K2 wash solution. Wait for the solution to flow through the column.
10	Apply 100 µl of K3 elution solution directly to the Genomic Mini AX column membrane. Wait for the eluate to flow through the column.
	Note. The purpose of this step is to decrease the total volume of eluate, since the column void volume is about 100 μ l.
11.	Transfer the Genomic Mini AX column to a new 2 ml tube (included).
	The column drop director possesses proper fitting that allows easy attachment to the precipitation tube.
12.	Add 1 ml of K3 elution solution. Wait for the eluate to flow through the column. Remove the Genomic Mini AX column.
13.	PM precipitation mix contains a precipitation enhancer and it should be intensively mixed before use by vigorous hand shaking.
	Add $800 \mu l$ of PM precipitation mix to the eluted DNA.
14.	Mix the sample by inverting the tube a few times and centrifuge for 10 min at 10 000 RPM .
	The light-blue DNA pellet should be visible at the bottom of the precipitation tube.
15.	Carefully discard supernatant. Be careful not to remove the DNA pellet at the bottom of the tube.
	Attention. When pouring out the supernatant, it is very easy to lose the DNA pellet. For safety, it is recommended to pour the supernatant into the prepared tube so the pellet can be recovered.
16.	Add 500 μl of 70% ethanol (not included). Mix the sample and centrifuge for 3 min at 10 000 RPM.
	Note. The light-blue DNA pellet should be visible at the bottom of the precipitation tube.
17.	Carefully discard supernatant. Be careful not to remove the DNA pellet at the bottom of the tube.
	Attention. When pouring out the supernatant, it is very easy to lose the DNA pellet. For safety, it is recommended to pour the supernatant into the prepared tube so the pellet can be recovered.
18.	Air dry the plasmid DNA pellet for 5 min at room temp . up-site down.
	Note. If there are any leftovers (small droplets) of alcohol on the tube walls they should be removed with sterile cotton buds.
19.	Dried DNA pellets can be dissolved in the desired volume of Tris buffer (included), TE buffer or sterile water (not included).
	Note. The blue color of DNA precipitate enables visual confirmation of the DNA dissolution process.

Store the DNA at -20 °C until later use.

Additional information

Pause in the purification process. The chromatography purification of DNA can be paused at any time while a sample is loaded onto a column. The purification process can be continued after a 15-hours-long pause with no influence on quality or quantity of purified DNA.

During the pause of the DNA purification the 15 ml tube with the column inside has to be closed with the screw cap to avoid membrane desiccation and subsequent DNA loss.

The volume of solution in the 15 ml tube enables the easy location of the procedure step after which the DNA purification process was paused:

- ~ 0.5 ml after the column equilibration;
- ~ 1.5 ml after the sample loading step;
- ~ 3 ml after the first washing step with K2 solution;
- ~ 4.5 ml after the second washing step with K2 solution.

Gravity flow of column. Genomic Mini AX column works by means of gravity. The flow rate strongly depends directly on the quantity and size of DNA molecules in a sample. High content of high molecular weight DNA decreases the flow rate. DNA amount exceeding 20 μ g loaded onto a column may lead to flow stoppage. In such cases the column should be centrifuged in a swing-out rotor for 1 min at 3000-4000 RPM. The centrifugation can be performed after the loading step and during the washing step with K2 solution or during the elution step with K3 solution.

Subsequently, the DNA elution step should be performed as follows:

Transfer the column to a new 15 ml tube (not included). Add 1 ml of K3 elution solution. Wait 2 min and centrifuge for 1 min at 3000 RPM. Transfer the eluate to a 2 ml tube (included). Follow point 13. of the protocol.

Safety Information

	Proteinase K
DANGER	H315 Causes skin irritation. H319 Causes serious eye irritation. H334 May cause allergy or asthma symptoms or breathing difficulties if inhaled. H335 May cause respiratory irritation. P261 Avoid breathing dust. P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,
	if present and easy to do. Continue rinsing. P342+P311 If experiencing respiratory symptoms call a Poison Center or doctor/physician.
	LS lysis suspension
	H302 Harmful if swallowed. H315 Causes skin irritation.
WARNING	H319 Causes serious eye irritation. P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
^	K1 equilibrating solution
WARNING	H302 Harmful if swallowed. H315 Causes skin irritation. H319 Causes serious eye irritation. P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
	K2 wash solution
	H225 Highly flammable liquid and vapor. H319 Causes serious eye irritation. H336 May cause drowsiness or dizziness. P210 Keep away from heat, sparks, open flames, hot surfaces. No smoking. P261 Avoid breathing vapors. P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
	K3 elution solution
	H225 Highly flammable liquid and vapor. H319 Causes serious eye irritation. H336 May cause drowsiness or dizziness. P210 Keep away from heat, sparks, open flames, hot surfaces. No smoking. P261 Avoid breathing vapors. P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
	PM precipitation mix
DANGER	H225 Highly flammable liquid and vapor. H319 Causes serious eye irritation. H336 May cause drowsiness or dizziness. P210 Keep away from heat, sparks, open flames, hot surfaces. No smoking. P261 Avoid breathing vapors. P305+P331+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.



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