A&A BIOTECHNOLOGY
innovating life science

Manual

MagnifiQ™ Genomic DNA reagents and consumables kit

A set of reagents and all necessary consumables for filling plates for the automated, magnetic isolation of DNA.

catalog #	size	compatible devices *
604D-16U-64	64 isolations	Auto-Pure 32A
604D-16V-64	64 isolations	Auto-Pure Mini
604D-16U-256	256 isolations	Auto-Pure 32A
604D-16V-256	256 isolations	Auto-Pure Mini
604D-96V-960	960 isolations	Auto-Pure 96

*** Compatible devices**
The kit has been tested with specific Allsheng brand isolation devices. This does not preclude it from working with other devices. If your device is not listed, please contact us at info@aabiot.com.

For research use only.

Guarantee
A&A Biotechnology provides a guarantee on this product.
The company does not guarantee the correct performance of this kit in the event of:

- not adhering to the supplied protocol
- use of not recommended equipment or materials
- use of other reagents than recommended or which are not a component of the product
- use of expired or improperly stored product or its components

Table of Contents

Advantages

- Automated, fast isolation.
- Enables isolation of different samples with universal kit and automated extraction programme.

Sample type

	sample size
Bacteria G-, G+ (cultures)	up to 2 x 10^8
Yeast (cultures)	up to 1 ml
Cell cultures	up to 1 x 10^6
Blood fresh or frozen, serum, plasma	up to 200 µl
Animal tissue	up to 20 mg
Swab	1 pc
Feces	20 - 50 mg
Feces (sample stored in conservation solution)	250 - 500 µl

Specification

protocol time	~ 30 min.
elution volume	50 – 100 µl
elution solution	Tris buffer
binding capacity	30 µg DNA
downstream applications	qPCR, RT-qPCR, sequencing

Description

MagnifiQ™ Genomic DNA reagents and consumables kit is designed for DNA isolation from various types of biological materials. Kit contains reagents and all necessary consumables for filling plates. The isolated material is perfect for further analyzes and tests by qPCR and RT-PCR methods and for sequencing.

The **MagnifiQ™** series products are based on the automated isolation of nucleic acids with use of magnetic beads. This method significantly shortens working time and reduces risk of mistake in comparison to manual methods.

Contents

Reagents

	64 isolations		256 isolations		960 isolations		
component	**quantity**	**cat #**	**quantity**	**cat #**	**quantity**	**cat #**	**storage**
MQBG binding mix	40 ml	K-MQBG-40	155 ml	K-MQBG-155	580 ml	K-MQBG-580	15–25 °C
A1WI wash solution	55 ml	K-A1WI-455	225 ml	K-A1WI-225	845 ml	K-A1WI-845	15–25 °C
Tris buffer	25 ml	K-TRIS-25	85 ml	K-TRIS-85	320 ml	K-TRIS-320	15–25 °C
LSDE buffer	35 ml	K-LSDE-35	140 ml	K-LSDE-140	530 ml	K-LSDE-530	15–25 °C
LTE 2X buffer	15 ml	K-LTE2X-15	55 ml	K-LTE2X-55	210 ml	K-LTE2X-210	15–25 °C
Proteinase K	3 ml	K-PRK-3	12 ml	K-PRK-12	42 ml	K-PRK-42	4–8 °C*

* Proteinase K can be stored at 15-25 °C for up to 12 months.

Plastic consumables

	604D-16U-64		604D-16U-256		
component	**quantity**	**cat #**	**quantity**	**cat #**	**storage**
2.2 ml plate	4 pcs	K-P96U22	16 pcs	K-P96U22	15–25 °C
tip comb 8	4 x 2 pcs	K-C8U-2	16 x 2 pcs	K-C8U-2	15–25 °C
protective film	4 pcs	K-MQF-4	16 pcs	K-MQF-16	15–25 °C

	604D-16V-64		604D-16V-256		
component	**quantity**	**cat #**	**quantity**	**cat #**	**storage**
2.2 ml plate	4 pcs	K-P96V22	16 pcs	K-P96V22	15–25 °C
tip comb 8	4 x 2 pcs	K-C8U-2	16 x 2 pcs	K-C8U-2	15–25 °C
protective film	4 pcs	K-MQF-4	16 pcs	K-MQF-16	15–25 °C

604D-96V-960

component	quantity	cat #	storage
CP - comb plate	1 pc	K-P96V22C	15 > 30 °C
2.2 ml plate	50 pcs	K-P96V22	15 > 30 °C
0.5 ml plate	2 x 5 pcs	K-P96V05-5	15 > 30 °C
tip comb 96	5 x 2 pcs	K-C96V-2	15 > 30 °C
protective film	10 pcs	K-MQF-10	15 > 30 °C

Additional equipment and reagents

Necessary

- automated pipette
- pipette tips
- 80% ethanol (1.6 ml per sample)

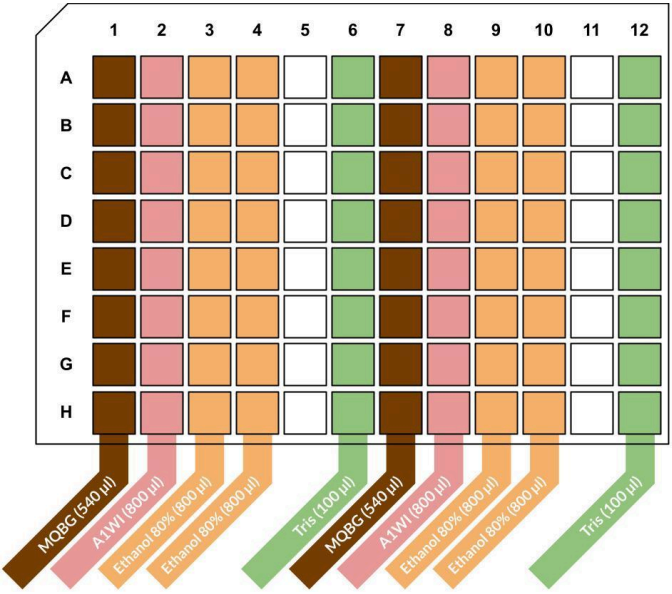
Optional

- RNAse (10 µl per sample), cat # 1006-10
- 1.5 ml Eppendorf tubes (sample lysis)
- 96 deep-well plates 2.2 ml (sample lysis)
- centrifuge with swing-out rotor for 96 deep-well plates
- vortex
- thermoblock
- protective film

Plate preparation

16 samples per plate format

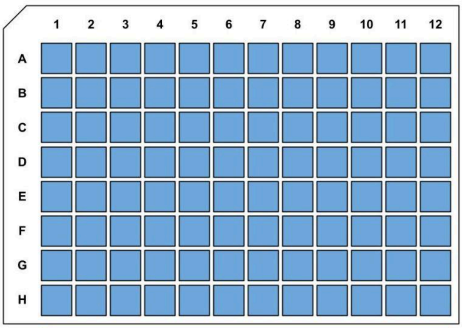
Distribute the buffers into a 2.2 ml plate as shown in the diagram below:



96 samples per plate format

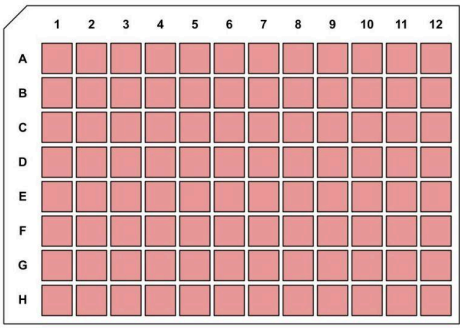
Distribute the buffers into a plates and mark them as shown in the diagram below:

"SP" (2.2 ml plate)



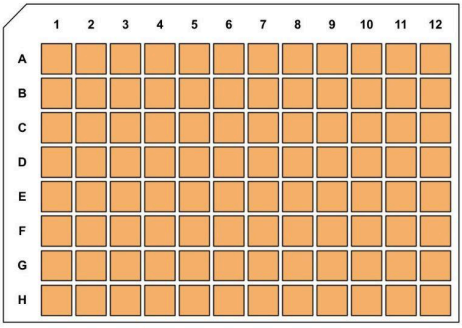
MQBG (540 µl)

"WP 1" (2.2 ml plate)



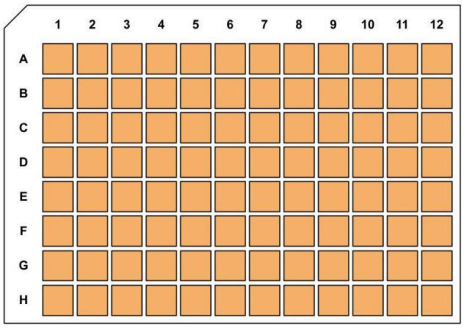
A1WI (800 µl)

"WP 2-3" (2.2 ml plate)



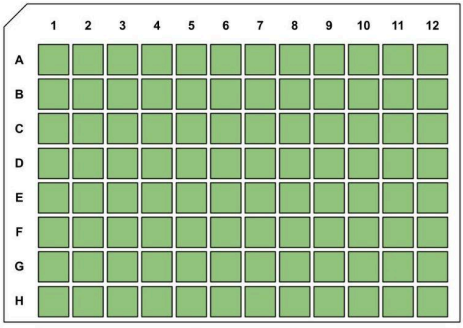
Ethanol 80% (800 µl)

"WP 2-3" (2.2 ml plate)



Ethanol 80% (800 µl)

"EP" (0.5 ml plate)



Tris (100 µl)

Material preparation

1.5 ml Eppendorf tubes

Bacteria G-, G+ (cultures)

Additional reagents you will need:

- **BacBreaker** bacteria lysis enzyme mix (20 µl per sample), cat # K-BACB-15A
- **BS** suspension buffer (200 µl per sample), cat # K-BS-30

Option:

- **Lysostaphin** (5 µl per sample), cat # 1007-3; For *Staphylococcus aureus* we recommend using lysostaphin.

1. Transfer the bacterial culture sample containing **2 x 10^8** bacteria to the 1.5 ml Eppendorf tube (not included).
   Centrifuge for **3 min** at **10 000 RPM**. Discard the supernatant.
2. Suspend the bacterial pellet in **200 µl** of **BS** buffer.
3. Add **20 µl BacBreaker** enzyme mix.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).

   Attention. For lysis of *Staphylococcus* bacteria, add **5 µl** of **lysostaphin**.
4. Vortex the sample for **10 s** and incubate for **10 min** at **50 °C**.

   Attention. For lysis of *Staphylococcus* bacteria with lysostaphin, mix and incubate for 10 min at 37 °C.
5. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K**.
6. Vortex the sample for **10 s** and incubate for **10 min** at **50 °C** with shaking.

   Note. If an automatic continuous shaking is not available, mix the samples by inverting the tubes a few times.
7. Centrifuge the sample for **2 min** at **10 000 RPM**.
8. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Yeast (cultures)

Additional reagents you will need:

- **Lyticase** (10 µl per sample), cat #1018-10
- **DTT RTU** (10 µl 1M solution per sample), cat # 2010-10P
- **BS** suspension buffer (200 µl per sample), cat # K-BS-30

Prepare **1M DTT** solution. Add 1 ml of sterile water (not included) to a vial containing DTT powder to obtain 1M DTT solution. Mix or vortex until complete dissolution of DTT powder. Store a clear solution at -20 °C.

1. Transfer **1 ml** of yeast culture to 1.5 ml Eppendorf tube (not included). Centrifuge for **3 min** at **10 000 RPM**. Discard the supernatant.
2. Suspend the yeast pellet in **200 µl** of **BS** buffer.
3. Add **10 µl lyticase** and **10 µl 1M DTT.**

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).
4. Vortex the sample for **10 s** and incubate for **15 min** at **37 °C**.
5. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K**.
6. Vortex the sample for **10 s** and incubate for **10 min** at **50 °C** with shaking.

   Note. If an automatic continuous shaking is not available, mix the samples by inverting the tubes a few times.
7. Centrifuge the sample for **2 min** at **10 000 RPM**.
8. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Cell cultures

1. Transfer the cell culture sample containing **1 x 10^6** cells to 1.5 ml Eppendorf tube (not included). Centrifuge for **3 min** at **10 000 RPM**. Discard the supernatant.
2. Suspend the cell pellet in **200 µl** of **Tris** buffer.
3. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K**.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).
4. Vortex the sample for **10 s** and incubate for **10 min** at **50 °C** with shaking.

   Note. If an automatic continuous shaking is not available, mix the samples by inverting the tubes a few times.
5. Follow point 1. of the protocol.

Blood: fresh or frozen, plasma, serum

1. Transfer **200 µl** the sample to 1.5 ml Eppendorf tube (not included).
2. Add **200 µl LTE 2X** and **20 µl Proteinase K**.
3. Vortex the sample for **10 s** and incubate for **10 min** at **50 °C** with shaking.

   Note. If automatic shaking is not available, mix the samples by inverting the tubes a few times.
4. Centrifuge for **20 s** at **10 000 RPM.**

   Note. Centrifuge to remove remaining material from lids of the tubes and deposit non-lysed material at the bottom of the tube.
5. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Animal tissue

1. Transfer **up to 20 mg** of fragmented animal tissue to 1.5 ml Eppendorf tube (not included).

   Note. The tissue should be fragmented by cutting into pieces or homogenization.

2. Add **400 µl LSDE** buffer and **40 µl Proteinase K.**

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).

3. Vortex the sample for **10 s** and incubate until complete lysis at **50 °C** with shaking.

   Note. If an automatic continuous shaking is not available, mix the samples by inverting the tubes a few times.

   Information. The lysis step can last from 1 to 12 hours. For maximum efficiency, lysis should be carried out until the tissue is completely dissolved in the lysis solution.

4. Centrifuge the sample for **2 min** at **10 000 RPM**.

5. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Swabs with transport medium

No additional material preparation is required.

Dry swabs

1. Break or cut off part of the swab with the collected sample and place it in a 1.5 ml Eppendorf tube (not included).

   Note. The portion of the swab with the collected sample should fit completely into the tube.

2. Add **500 µl** of **LSDE** buffer and **20 µl Proteinase K.**

   Note. Part of the swab with the sample should be completely immersed in the buffer.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).

3. Vortex the sample for **10 s** and incubate for **10 min** at **50 °C** with shaking.

   Note. If an automatic continuous shaking is not available, mix the samples by inverting the tubes a few times.

4. **Attention.** For the isolation process, take the entire volume of the sample, but not more than 400 µl.

   Follow point 1. of the protocol.

Feces (microbiome including G+, G- bacteria)

Additional reagents you will need:

- **Bead-beat tubes** (2 ml screwed Bead-beat tubes containing beads mix), cat # K-PKCM-50
- **L3P** precipitation solution (100 µl per sample), cat # K-L3P-60
- **LSDE** buffer (additional 500 µl per sample), cat # K-LSDE-500
- **antifoam** (10 µl per sample), cat# K-AYS-1

Before starting the process, mix the **LSDE** buffer with **antifoam**. Prepare the **LSDE-antifoam** mix by combining **1 ml** of **LSDE** buffer with **10 µl** of **antifoam** per sample. Prepare a volume sufficient for the number of isolated samples with a 10% excess. Mix well before use.

1. Transfer **20-50 mg** of stool sample to the 2 ml screwed Bead-beat tube containing beads mix.
   Add **1 ml** of **LSDE-antifoam** buffer.

2. **Option A:** Bead Beating. Use the suitable tube bead beater with the following programme:
   3 x run of 20 s at **maximum force** with **2 min** cool down rest.

   Option B: Place the tubes in the incubator with mixing function (e.g. Thermomixer) and vortex at **2000 RPM** for **30 min** at **room temperature**.

3. Centrifuge for **5 min** at **10 000 RPM**.

4. Transfer **500 µl** of supernatant to a new 1.5 ml Eppendorf tube (not included).

5. Add **20 µl Proteinase K.**

6. Vortex the sample for **10 s** and incubate for **15 min** at **50 °C** with shaking **1400 RPM**.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10) and incubate for **10 min** at **37 °C** with shaking at **1400 RPM.**

7. Add **100 µl** of **L3P** precipitation solution. Close the tube and mix whole content by inverting the tube.

8. Place on ice for **3 min.**

9. Centrifuge the sample for **5 min** at **10 000 RPM**.

10. **Attention.** In the isolation protocol, use the supernatant as the sample.

    Follow point 1. of the protocol.

Feces (sample stored in conservation solution StoolSave™ DNA Protection kit)

Additional reagents you will need:

For stool samples stored in the **StoolSave™ DNA Protection kit (cat # 006-10)**:

- **Bead-beat tubes** (2 ml screwed Bead-beat tubes containing beads mix), cat # K-PKCM-50
- **L3P** precipitation solution (100 µl per sample), cat # K-L3P-60

For stool samples stored in another preservation solution:

- **Bead-beat tubes** (2 ml screwed Bead-beat tubes containing beads mix), cat # K-PKCM-50
- **L3P** precipitation solution (100 µl per sample), cat # K-L3P-60
- **LSDE** buffer (additional 250 µl per sample),cat # K-LSDE-500
- **antifoam** (10 µl per sample), cat# K-AYS-1

1. **Feces stored in conservation solution StoolSave™ DNA Protection kit**:

   Transfer **500 µl** of stool sample suspended in conservation/ transportation solution to the 2 ml screwed Bead-beat tube containing zirconia beads.
   Add **500 µl** of **LSDE** buffer.

   Feces stored in another preservation solution:

   Before starting the process, mix the **LSDE** buffer with **antifoam**. Prepare the **LSDE-antifoam** mix by combining **750 µl** of **LSDE** buffer with **10 µl** of **antifoam** per sample. Prepare a volume sufficient for the number of isolated samples with a 10% excess. Mix well before use.

   Transfer **250 µl** of stool sample suspended in conservation/ transportation solution to the 2 ml screwed Bead-beat tube containing zirconia beads.
   Add **750 µl** of **LSDE-antifoam** buffer.

2. **Option A:** Bead Beating. Use the suitable tube bead beater with the following programme:
   3 x run of 20 sec at **maximum force** with **1 min** cool down.

   Option B: Place the tubes in the incubator with mixing function (e.g. Thermomixer) and vortex at **2000 RPM** for **30 min** at **room temperature**.

3. Centrifuge for **5 min** at **10 000 RPM**.

4. Transfer **500 µl** of supernatant to a new 1.5 ml Eppendorf tube (not included).

5. Add **20 µl Proteinase K.**

6. Vortex the sample for **10 s** and incubate for **15 min** at **50 °C** with shaking **1400 RPM**.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10) and incubate for **10 min** at **37 °C** with shaking at **1400 RPM.**

7. Add **100 µl** of **L3P** precipitation solution. Close the tube and mix whole content by inverting the tube.

8. Place on ice for **3 min.**

9. Centrifuge the sample for **5 min** at **10 000 RPM**.

10. **Attention.** In the isolation protocol, use the supernatant as the sample.

Follow point 1. of the protocol.

96 deep-well plates 2.2 ml

Bacteria G-, G+ (cultures)

Additional reagents you will need:

- **EMXB** bacteria lysis enzyme mix (20 µl per sample), cat # K-EMXB-15A
- **BS** suspension buffer (200 µl per sample), cat # K-BS-250

Option:

- **Lysostaphin** (5 µl per sample), cat # 1007-3; For *Staphylococcus aureus* we recommend using lysostaphin.

1. Transfer the bacterial culture samples containing **2 x 10^8** bacteria to the 96 deep-well plate (not included).
   Seal the plate with a protective film and centrifuge for **10 min** at **1000 x g**.
   Remove the protective film.
   Carefully discard the supernatant with a pipette.
2. Suspend the bacterial pellet in **200 µl** of **BS** buffer.
3. Add **20 µl EMXB** enzyme mix to the wells.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).
   Attention. For lysis of *Staphylococcus* bacteria, add **5 µl** of **lysostaphin**.
4. Mix the contents of the wells by pipetting.
   Seal the plate with a protective film and incubate for **20 min** at **55 °C** with shaking **1600 RPM.**
   Remove the protective film.

   Attention. For lysis of *Staphylococcus* bacteria with lysostaphin, mix and incubate for 20 min at 42°C.
5. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K** to the wells.
   Mix the contents of the wells by pipetting.
6. Seal the plate with a protective film and incubate for **20 min** at **55 °C** with shaking **1600 RPM.**
7. Centrifuge the sample for **10 min** at **1000 x g**.
8. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Yeast (cultures)

Additional reagents you will need:

- **Liticase** (10 µl per sample), cat #1018-10
- **DTT RTU** (10 µl 1M solution per sample), cat # 2010-10P
- **BS** suspension buffer (200 µl per sample), cat # K-BS-250

Prepare **1M DTT** solution. Add 1 ml of sterile water (not included) to a vial containing DTT powder to obtain 1M DTT solution. Mix or vortex until complete dissolution of DTT powder. Store a clear solution at -20 °C.

1. Transfer **1 ml** of yeast culture to the 96 deep-well plate (not included).
Seal the plate with a protective film and centrifuge for **10 min** at **1000 x g**.
Remove the protective film.
Carefully discard the supernatant with a pipette.

2. Suspend the yeast pellet in **200 µl** of **BS** buffer.

3. Add **10 µl lyticase** and **10 µl 1M DTT** to the wells.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).

4. Mix the contents of the wells by pipetting.
Seal the plate with a protective film and incubate for **20 min** at **37°C** with shaking **1600 RPM.**
Remove the protective film.

5. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K** to the wells.
Mix the contents of the wells by pipetting.

6. Seal the plate with a protective film and incubate for **20 min** at **55 °C** with shaking **1600 RPM.**

7. Centrifuge the sample for **10 min** at **1000 x g**.

8. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Cell cultures

1. Transfer the cell culture samples containing **1 x 10^6** cells to the 96 deep-well plate (not included).
   Seal the plate with a protective film and centrifuge for **10 min** at **1000 x g**.
   Remove the protective film.
   Discard the supernatant with pipette.
2. Suspend the cell pellet in **200 µl** of **Tris** buffer.
3. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K** to the wells.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).
4. Mix the contents of the wells by pipetting.
   Seal the plate with a protective film and incubate for **20 min** at **55 °C** with shaking **1600 RPM.**
5. Follow point 1. of the protocol.

Blood: fresh or frozen, plasma, serum

1. Transfer **200 µl** the sample to the 96 deep-well plate (not included).
2. Add **200 µl LTE 2X** buffer and **20 µl Proteinase K** to the wells.
   Mix the contents of the wells by pipetting.
3. Seal the plate with a protective film and incubate for **20 min** at **55 °C** with shaking **1600 RPM.**
4. Centrifuge for **1 min** at **1000 x g**.

   Note. Centrifuge to remove remaining material from lids of the tubes and placement non-lysed material at the bottom of the plate.
5. Follow point 1. of the protocol.

Animal tissue

1. Transfer **up to 20 mg** of fragmented animal tissue to the 96 deep-well plate (not included).

   Note. The tissue should be fragmented by cutting into pieces or homogenization.

2. Add **400 µl LSDE** buffer and **40 µl Proteinase K** to the wells.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).

3. Mix the contents of the wells by pipetting.
   Seal the plate with a protective film and incubate until complete lysis at **55 °C** with shaking **1600 RPM.**

   Information. The lysis step can last from 1 to 12 hours. For maximum efficiency, lysis should be carried out until the tissue is completely dissolved in the lysis solution.

4. Centrifuge for **10 min** at **1000 x g**.

5. **Attention.** In the isolation protocol, use the supernatant as the sample.

   Follow point 1. of the protocol.

Swabs with transport medium

No additional material preparation is required.

Dry swabs

1. Break or cut off part of the swab with the collected sample and place it in the wells of 96 deep-well plate (not included).

2. Add **500 µl** of **LSDE** buffer and **20 µl Proteinase K** to the wells.

   Note. Part of the swab with the sample should be completely immersed in the buffer.

   Optional RNA removal. Add **10 µl** of **RNAse** (cat # 1006-10).

3. Mix the contents of the wells by pipetting.
   Seal the plate with a protective film and incubate for **20 min** at **55 °C** with shaking **1600 RPM.**

4. **Attention.** For the isolation process, take the entire volume of the sample, but no more than 400 µl.

   Follow point 1. of the protocol.

Protocol

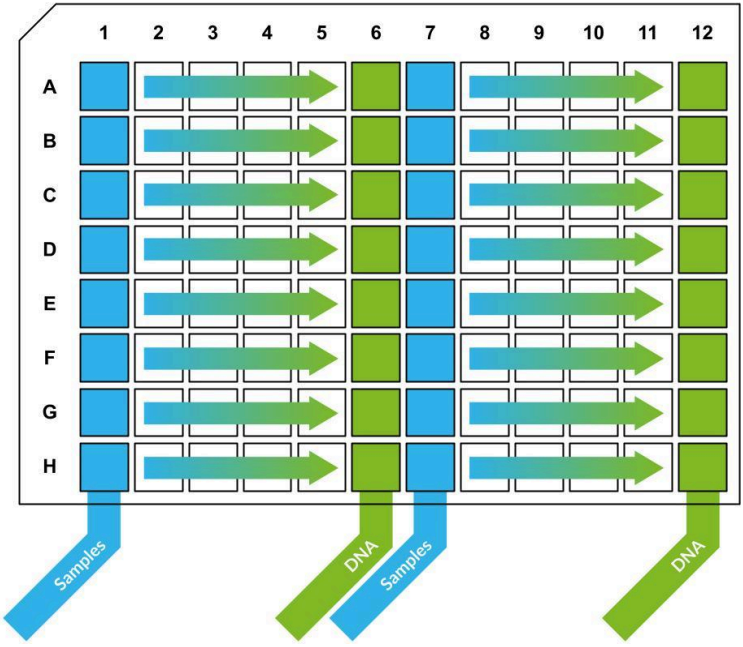
Protocol files

device	protocol name	protocol file	installation
Auto-Pure Mini	MQ-UND-MI	aabiot.com/protocols/magnifiq/MI/MQ-UND-MI.txt	1. Create folder "items" on a USB drive and copy the protocol file to it. 2. Insert the USB drive into a USB slot in the device. 3. On a device screen, go to Settings > System > Transfer >Import. 4. Select the protocol and tap "Import".
Auto-Pure Mini (QR code)	MQ-UND-MI		1. On a device screen, go to Run > ☰ > ⌜⌝ 2. Scan the QR code with the device's scanner.
Auto-Pure 32A	MQ-UND-32A	aabiot.com/protocols/magnifiq/32A/MQ-UND-32A.txt	1. Create folder "items" on a USB drive and copy the protocol file to it. 2. Insert the USB drive into a USB slot in the device. 3. On a device screen, go to Settings > Im.&Export > Import. 4. Select the protocol and tap "Import."
Auto-Pure 96	MQ-UND-96	aabiot.com/protocols/magnifiq/96/MQ-UND-96.txt	
Auto-Pure S32	MQ_UND_S32	aabiot.com/protocols/magnifiq/S32/MQ_UND_S32.txt	1. Create folder "im_export_protocols" on a USB drive and copy the protocol file to it. 2. Insert the USB drive into a USB slot in the device. 3. On a device screen, go to Protocols >Import. 4. Select the protocol and tap "Import".

16 samples per plate format

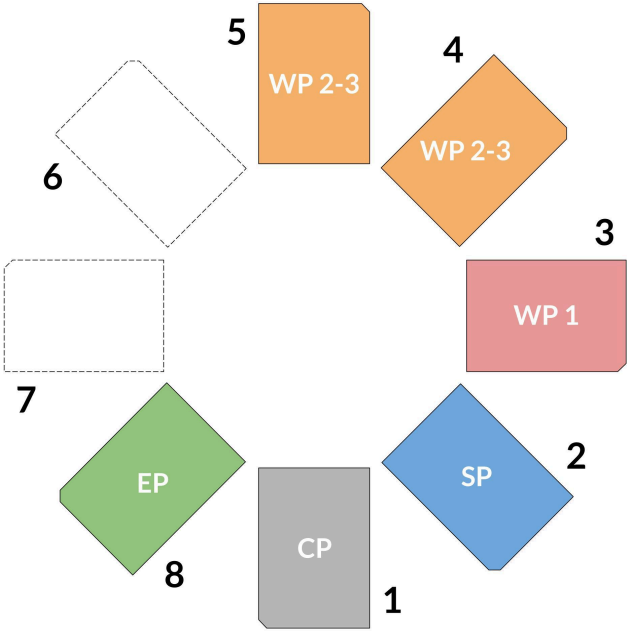
1. Add **400 µl** of samples to the wells in columns **1** and **7** of **2.2 ml plate**.
2. Place one or two **2.2 ml plates** in the extraction device.
3. Place the appropriate number of **tip combs 8** in the extraction device.
4. Run the protocol on your device.
5. After the program is over, remove the combs and then remove the **2.2 ml plate** from the extraction device and seal it with **protective film**. The extracted DNA is located in columns **6** and **12**.

Note. For longer storage of extracted material, transfer it from the plate to appropriate tubes and store at 4 °C.



96 samples per plate format

1. Add **400 µl** of sample to the wells of the **SP** plate.
2. Place the **tip comb 96** into the **CP** plate.
3. Place the plates on the working table of the extraction device according to the diagram below:



4. Run the protocol on your device.
5. After the program is over, remove the **EP plate** from the extraction device and seal it with a **protective film.**

   Note. For longer storage of extracted material, transfer it from the plate to appropriate tubes and store at 4 °C.
6. Discard remaining plates except the **CP** plate, which can be reused.

Safety information

DANGER

Proteinase K

H315 Causes skin irritation.
H319 Causes serious eye irritation.
H334 May cause allergy or asthma symptoms or breathing difficulties if inhaled.
H335 May cause respiratory irritation.
P261 Avoid breathing dust.
P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
P342+P311 If experiencing respiratory symptoms call a Poison Center or doctor/physician.

WARNING

LTE 2X

H302 Harmful if swallowed.
H315 Causes skin irritation.
H319 Causes serious eye irritation.
P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

DANGER

MQBG binding mix

H302+H312+H332 Harmful if swallowed, in contact with skin or if inhaled.
H314 Causes severe skin burns and eye damage.
H412 Harmful to aquatic life with long lasting effects.
P273 Avoid release to the environment.
P280 Wear protective gloves/protective clothing/eye protection/face protection/hearing protection.
P301+P312+P330 If swallowed: Call a poison center/doctor/ if you feel unwell.
P303+P361+P353 If on skin (or hair): Take off immediately all contaminated clothing. Rinse skin with water or shower.
P304+P340 If inhaled: Remove person to fresh air and keep comfortable for breathing.
P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

DANGER

A1WI wash solution

H225 Highly flammable liquid and vapor.
H302 Harmful if swallowed.
H315 Causes skin irritation.
H319 Causes serious eye irritation.
H336 May cause drowsiness or dizziness.
P210 Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking.
P261 Avoid breathing dust/fume/gas/mist/vapours/ spray.
P280 Wear protective gloves/protective clothing/eye protection/face protection/hearing protection.
P301+P312+P330 If swallowed: Call a poison center/doctor/ if you feel unwell.
P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.
P337+P313 If eye irritation persists: Get medical advice/ attention.

A&A BIOTECHNOLOGY

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